

## Notes

## Novel Inhibitors for Multidrug Resistance: 1,3,5-Triazacycloheptanes

Hiroyuki Sawanishi,<sup>†</sup> Shinya Wakusawa,<sup>\*†</sup> Rieko Murakami,<sup>†</sup> Hiromi Muramatsu,<sup>†</sup> Hirokazu Suzuki,<sup>†</sup> Akemi Takashima (née Sano),<sup>†</sup> Tatsuo Aizawa,<sup>‡</sup> and Ken-ichi Miyamoto<sup>†</sup>

Faculty of Pharmaceutical Sciences, Hokuriku University, Ho-3, Kanagawa-machi, Kanazawa 920-11, Japan, and Tsumura & Company, 2 Rokubancho, Chiyoda-ku, Tokyo 102, Japan

Received May 30, 1995<sup>⊗</sup>

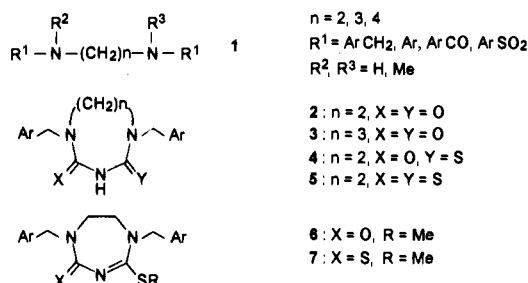
1,3,5-Triazacycloheptanes were synthesized and examined for reversal of the multidrug resistance dependent on P-glycoprotein. Most of these compounds increased the intracellular uptake of vinblastine in multidrug-resistant mouse leukemia P388/ADR cells without influence upon the vinblastine accumulation in P388/S cells. The efficacy of 1,5-dibenzyl-1,3,5-triazacycloheptanes in increasing the vinblastine accumulation was in the order of 2,4-dithioxo (**5**) > 2-oxo-4-thioxo (**4**) ≈ 4-(methylthio)-2-oxo (**6**) > 2,4-dioxo (**2**). The efficacy was further increased when the benzyl group was converted to a chlorobenzyl group. Among these compounds, **6c** [1,5-bis(4-chlorobenzyl)-1,5,6,7-tetrahydro-4-(methylthio)-2H-1,3,5-triazepin-2-one] potentiated the *in vitro* cell growth-inhibitory effect of vinblastine, adriamycin, and mitomycin C on P388/ADR cells and prolonged the life span of P388/ADR-bearing mice in combined therapy with vinblastine more than vinblastine alone.

## Introduction

Multidrug resistance (MDR) is a major obstacle in cancer chemotherapy.<sup>1,2</sup> Multidrug-resistant cells generally show resistance to antitumor agents of natural products such as Vinca alkaloids and anthracyclines. Classical multidrug-resistant cells express an outward drug transporter, P-glycoprotein, in the plasma membrane. This protein increases the efflux of antitumor drugs, and the intracellular concentration of these drugs is kept at a low level.

Therefore, the reversal of MDR is a problem that must be overcome for the progress of cancer chemotherapy. There are several efforts to reverse MDR by nonantitumor agents.<sup>3-9</sup> Most of these drugs share a common structural feature of aromatic rings and amino groups located between the rings. These structural features seemed to be involved in counteracting the action of P-glycoprotein. In a previous study,<sup>10</sup> we synthesized a series of diamines, dicarboxamides, and disulfonamides with various aryl groups at both termini (**1**) and showed that *N*-methylethylenediamine and *N*-methyl-ethylenedisulfonamides with terminal methyl- or chloro-substituted benzene rings showed potent efficacy on vinblastine (VBL) uptake in multidrug-resistant P388/ADR cells that overexpress P-glycoprotein.<sup>11</sup> Moreover, these compounds as well as reported MDR inhibitors have inadequate *in vivo* effects for clinical use. On the basis of these observations, we designed medium-sized azacycloalkane compounds to reverse MDR. This study deals with synthesis and structure-activity relationships of 1,3,5-triazacycloalkane derivatives **2-7** on VBL accumulation of P388 cells and examined the inhibitory effects on P388/ADR cells *in vitro* and *in vivo*.

## Chart 1



## Chemistry

Hagemann<sup>12</sup> has reported the synthesis of 1,5-dibenzyl-1,3,5-triazacycloheptane-2,4-dione (**2a**) from *N,N'*-dibenzylethylenediamine **8a** and dimethyl imidodicarboxylate. We prepared 1,3,5-triazacycloheptane-2,4-diones **2b-d** and the new 1,3,5-triazacyclooctane-2,4-diones **3a-d** by the reaction of ethylenediamine **8** or trimethylenediamine **9** with diethyl imidodicarboxylate by the procedure of Hagemann (Scheme 1). **2** reacted with 2,4-bis(4-methoxyphenyl)-1,3-dithia-2,4-diphosphetane 2,4-disulfide (Lawesson's reagent) to give a mixture of the 2-oxo-4-thioxo **4** and the 2,4-dithioxo **5**. Treatment of **4** or **5a** with methyl iodide in the presence of sodium hydride, respectively, afforded the methylthio oxo **6** or the methylthio thioxo **7a**.

## Pharmacological Results and Discussion

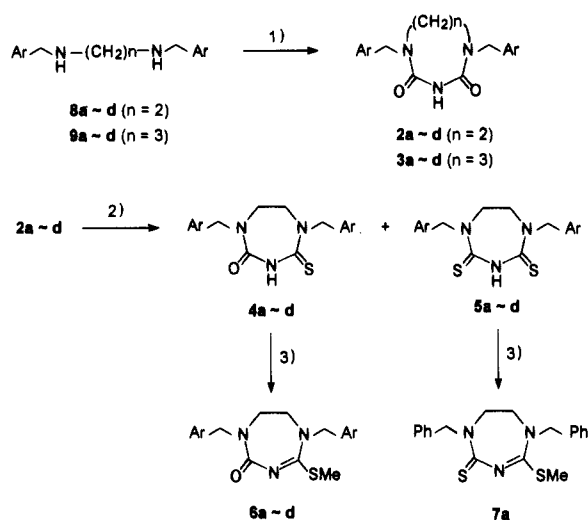
The effects of a series of 1,3,5-triazacycloalkanes (**2-7a**) on the VBL uptake in drug-sensitive P388 cells (P388/S) and multidrug-resistant P388 cells (P388/ADR) are shown in Table 1. P388/ADR cells intrinsically accumulated VBL about 4-fold less and are resistant to VBL about 40-fold more than P388/S cells. These compounds at 10  $\mu$ M hardly or only slightly increased the VBL accumulation in P388/S cells, but most of compounds promoted the VBL accumulation in P388/

\* To whom correspondence should be addressed.

<sup>†</sup> Hokuriku University.

<sup>‡</sup> Tsumura & Co.

<sup>⊗</sup> Abstract published in *Advance ACS Abstracts*, October 15, 1995.

Scheme 1<sup>a</sup>

<sup>a</sup> (1)  $\text{HN(CO}_2\text{Et)}_2$ , (2) Lawesson's reagent, (3) NaH, MeI.

**Table 1.** Effects of Triazacycloalkane Derivatives on Accumulation of VBL in P388/S and P388/ADR Cells

compd	relative uptake (-fold) <sup>a</sup>		compd	relative uptake (-fold) <sup>a</sup>	
	P388/S	P388/ADR		P388/S	P388/ADR
2a	1.09	1.41	5a	1.26	4.16
2b	1.20	1.05	5b	1.34	5.34
2c	1.22	3.44	5c	1.48	6.00
2d	0.93	1.16	5d	1.21	5.26
3a	0.93	1.78	6a	1.37	2.13
3b	1.12	2.62	6b	1.16	6.75
3c	1.17	2.86	6c	1.28	6.87
3d	1.06	1.11	6d	1.04	1.14
4a	1.20	2.11	7a	1.19	3.44
4b	1.26	6.14	8c	1.04	2.62
4c	1.30	5.35	verapamil	1.02	5.70
4d	1.14	2.36			

<sup>a</sup> Relative VBL uptake (-fold) in drug (10  $\mu\text{M}$ )-treated cells to the uptake in untreated cells. The uptake of [<sup>3</sup>H]VBL in untreated P388/S and P388/ADR cells was  $30\,400 \pm 758.2$  and  $7814 \pm 520.4$  dpm/ $10^6$  cells, respectively.

ADR cells. In P388/ADR cells, comparing the efficacy of the dioxo compounds **2a-d** and **3a-d**, the compounds with same side chain were almost equipotent without distinction of the ring size of  $n = 2$  or 3, and **2c** was approximately equipotent to **8c**, a starting material (Scheme 1). In addition, among the compounds with benzyl groups (**2-6a**) or 3,4-dimethoxybenzyl groups (**2-6d**), the efficacy of the compounds with the same triazacycloalkane ring was equipotent, and the efficacy was in the order of dithio (**5**) > oxo thio (**4**)  $\approx$  methylthio oxo (**6**) > dioxo (**2, 3**). The methylthio thio compound **7a** was almost equipotent to the methylthio oxo compound **6a**. On the other hand, substitution on the 3- or 4-chlorobenzyl group increased the efficacy as in **2-6b,c**, and the substitution effect was dramatic in the cases of **4** and **6**. **4b** and **6b,c** increased the VBL accumulation 6-fold in P388/ADR cells. The effects of these compounds were almost equipotent to that of verapamil, and the effect of **6c** was much more potent than that of **8c** (Table 1).

The action of the most effective compound, **6c**, was further investigated as follows. *In vitro* 50% cell growth-inhibitory concentrations of **6c** against P388/S and P388/ADR cells were 13 and 18  $\mu\text{M}$ , respectively. Noncytotoxic concentrations of **6c** potentiated the growth-inhibitory effect of VBL on P388/ADR cells in a dose-dependent manner, while this compound affected the

**Table 2.** *In Vitro* Combination Effects of **6c** with VBL in P388/S and P388/ADR Cells<sup>a</sup>

6c ( $\mu\text{M}$ )	IC <sub>50</sub> of VBL (nM)	
	P388/S	P388/ADR
0	1.7 (1.0)	70(1.0)
1.0	0.9 (1.9)	23(3.2)
2.5	0.7 (2.4)	6.2 (11)
5.0	0.4 (4.3)	2.2 (23)

<sup>a</sup> Cells were cultured with or without VBL in the presence or absence of **6c** for 48 h. Numbers in parentheses represent the relative increase in the growth-inhibitory effect of VBL by the combined treatment with **6c**.

**Table 3.** *In Vitro* Combination Effects of **6c** with Adriamycin, Mitomycin C, and Cisplatin in P388/S and P388/ADR Cells<sup>a</sup>

antitumor drug	IC <sub>50</sub> of antitumor drugs (nM)			
	P388/S		P388/ADR	
	-	+	-	+
adriamycin	19	13	360	90
mitomycin C	48	44	535	264
cisplatin	48	72	61	51

<sup>a</sup> Cells were cultured in the absence or presence of various concentrations of an antitumor agent and 2.5  $\mu\text{M}$  **6c** for 48 h. The sign of - or + indicates the IC<sub>50</sub> values of antitumor agent alone or those of combined treatment with **6c** and antitumor agent, respectively.

**Table 4.** Antitumor Combination Therapy with VBL and **6c** in P388/ADR-Bearing Mice<sup>a</sup>

cell	treatment	survival days (average $\pm$ SD)	ILS (%)
P388/ADR	none	9.6 $\pm$ 0.5	0
	VBL (0.2 mg/kg)	11.0 $\pm$ 0.6	14.6
	VBL + <b>6c</b> (10 mg/kg)	11.6 $\pm$ 0.5*	20.8
	VBL + <b>6c</b> (20 mg/kg)	13.2 $\pm$ 1.5*	37.5
	VBL + <b>6c</b> (40 mg/kg)	14.0 $\pm$ 1.7*	45.8
P388/S	none	10.0 $\pm$ 0.9	4.2
	VBL (0.2 mg/kg)	8.2 $\pm$ 0.8	0
	VBL (0.2 mg/kg)	15.5 $\pm$ 1.4*	89.0

<sup>a</sup> Each mouse was implanted with  $1 \times 10^6$  cells on day 0 and treated with intraperitoneal injection of VBL and/or oral administration of **6c** on days 1-4 and 6-8. \* $P < 0.05$  (Student's *t*-test or two-sample test with Welch's correction) compared to the no treatment group.

effect of VBL on P388/S cells only a little (Table 2). In P388/ADR cells, most of the potentiation of the effect of VBL by **6c** is probably caused on the increase in VBL uptake, although other unknown mechanisms may be partly involved in the potentiation since smaller potentiation was seen in P388/S cells. When **6c** at 2.5  $\mu\text{M}$  was combined with adriamycin, mitomycin C, or cisplatin, it potentiated the effects of adriamycin and mitomycin C 4- and 2.5-fold, respectively, in P388/ADR cells without influence on the effect of cisplatin. In P388/S cells, **6c** did not affect the effects of these antitumor drugs. Consequently, **6c** selectively potentiated the effects of MDR-related drugs in the resistant cells (Table 3).

Next, we examined the combined effects of **6c** with VBL on P388/ADR-bearing mice. As shown in Table 4, oral administration of **6c** alone did not increase the survival days of the mice, and intraperitoneal injection of VBL at 0.2 mg/kg prolonged the life span only 15%, while it was prolonged 89% in P388/S-bearing mice. When VBL was combined with **6c** in P388/ADR-bearing mice, the life span was dose-dependently elongated by **6c**.

P388/ADR cells were shown to express P-glycoprotein in the plasma membrane and to extrude it in a energy-

dependent manner.<sup>11</sup> Since **6c** and the related compounds showed more potent action in P388/ADR cells than in P388/S cells negative for P-glycoprotein, these compounds are thought to have their antiresistant effect through an inhibitory action on P-glycoprotein.

In this study, the effects of triazacycloalkanes with an aryl side chain were investigated on the anti-MDR effect *in vitro* and *in vivo*. The effects of these compounds on VBL uptake seem to be due to the triazacycloalkane ring structures because the efficacy of the dibenzyl compounds was in the order of dithioxo (**5**) > oxo thioxo (**4**) ≈ methylthio oxo (**6**) > dioxo (**2**, **3**). These may indicate that the thioxo or methylthio group containing sulfur atoms could be better for the activity, as indicated in our previous reports that isoquinoline-sulfonamides<sup>13</sup> and ethylenedisulfonamides<sup>10</sup> are effective *in vitro* inhibitors of MDR. On the other hand, the efficacy was inversely related to the number of the oxo groups in 1,3,5-triazacycloalkanes. This may be similar to our previous evidence that the activity of dicarboxamide compounds was less than that of diamines.<sup>10</sup> The conversion of benzyl or the 3,4-dimethoxybenzyl group into a 3- or 4-chlorobenzyl group increased the efficacy of the oxo thioxo (**4**) or methylthio oxo (**6**) compounds for VBL accumulation in P388/ADR cells. These effects may be due to a moderate increase in the lipid solubility or in the affinity for a target molecule P-glycoprotein.

In conclusion, from the structure-activity relationships of 1,3,5-triazacyclohexanes and -octanes for VBL accumulation in P388/ADR cells, we found the bis(4-chlorobenzyl) derivative **6c** to be an effective inhibitor of MDR.

## Experimental Section

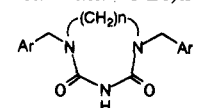
**General.** All melting points were measured on a Yanagimoto micromelting point hot stage apparatus and are uncorrected. Infrared (IR) spectra were taken with a Hitachi 270-30 spectrometer, and mass spectra (MS) were measured with a JEOL DX-300 instrument. <sup>1</sup>H-NMR spectra were recorded on a JEOL PMX-60 spectrometer in CDCl<sub>3</sub> using tetramethylsilane as an internal standard; spectral assignments were confirmed by spin-decoupling experiments. Experimental analyses were done with a Perkin-Elmer 240C elemental analysis apparatus on solid samples only; the analytical results (C, H, N) are within ±0.4% of the theoretical values.

**Starting Materials.** *N,N'*-Bis(4-chlorobenzyl)ethylenediamine (**8**),<sup>14</sup> *N,N'*-bis(3,4-dimethoxybenzyl)ethylenediamine (**8d**),<sup>15</sup> *N,N'*-dibenzyltrimethylenediamine (**9a**),<sup>16</sup> *N,N'*-bis(3-chlorobenzyl)trimethylenediamine (**9b**),<sup>17</sup> *N,N'*-bis(4-chlorobenzyl)trimethylenediamine (**9c**),<sup>18</sup> and *N,N'*-bis(3,4-dimethoxybenzyl)trimethylenediamine (**9d**)<sup>19</sup> were prepared by the reported methods.

***N,N'*-Bis(3-chlorobenzyl)ethylenediamine (8b).** 3-Chlorobenzaldehyde (15.17 g, 108 mmol) was added to a stirred solution of ethylenediamine (3.24 g, 54 mmol) in MeOH (20 mL) at 0–5 °C. The reaction mixture was stirred for 3 h at room temperature, and the imino compound precipitated as a solid was collected by filtration. A mixture of the imino compound, sodium borohydride (6.78 g), and MeOH (150 mL) was refluxed for 3 h. The solvent was then evaporated, and water was added to the residue. The aqueous mixture was extracted with benzene, and the extracts were washed with water, dried, and evaporated *in vacuo*. The residue was chromatographed on silica gel using Et<sub>2</sub>O:MeOH (95:5) as eluent to give **8b**: 7.35 g, 95% yield; viscous oil; IR (KBr) 3316 cm<sup>-1</sup> (NH); <sup>1</sup>H-NMR δ 1.59 (2H, br s), 2.73 (4H, s), 3.74 (4H, s), 7.10–7.51 (8H, m); MS *m/z* 308 (M<sup>+</sup>); dihydrochloride (C<sub>16</sub>H<sub>8</sub>N<sub>2</sub>Cl<sub>2</sub>·2HCl) mp 300 °C dec. Anal. (C<sub>16</sub>H<sub>10</sub>N<sub>2</sub>Cl<sub>4</sub>) C, H, N.

**1,5-Bis(3-chlorobenzyl)-1,3,5-triazacycloheptane-2,4-dione (2b).** A solution of *N,N'*-bis(3-chlorobenzyl)ethylenediamine (**8b**) (1.55 g, 5 mmol) and diethyl iminodicyoxylate

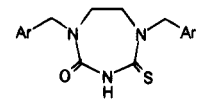
**Table 5.** Physicochemical Data for **2c,d** and **3a–d**



compd	<i>n</i>	Ar	yield (%)	mp (°C) <sup>a</sup>	formula <sup>b</sup>
<b>2c</b>	2	4-ClC <sub>6</sub> H <sub>4</sub>	53	158–159	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> O <sub>2</sub> Cl <sub>2</sub>
<b>2d</b>	2	3,4-(MeO) <sub>2</sub> C <sub>6</sub> H <sub>3</sub>	60	136–137	C <sub>23</sub> H <sub>27</sub> N <sub>3</sub> O <sub>6</sub>
<b>3a</b>	3	C <sub>6</sub> H <sub>5</sub>	66	177–179	C <sub>19</sub> H <sub>21</sub> N <sub>3</sub> O <sub>2</sub>
<b>3b</b>	3	3-ClC <sub>6</sub> H <sub>4</sub>	46	148–149	C <sub>19</sub> H <sub>19</sub> N <sub>3</sub> O <sub>2</sub> Cl <sub>2</sub>
<b>3c</b>	3	4-ClC <sub>6</sub> H <sub>4</sub>	30	197–198	C <sub>19</sub> H <sub>19</sub> N <sub>3</sub> O <sub>2</sub> Cl <sub>2</sub>
<b>3d</b>	3	3,4-(MeO) <sub>2</sub> C <sub>6</sub> H <sub>3</sub>	48	141–143	C <sub>23</sub> H <sub>29</sub> N <sub>3</sub> O <sub>6</sub>

<sup>a</sup> All compounds were recrystallized from ethyl acetate. <sup>b</sup> C, H, and N analyses are within ±0.4% of theoretical values.

**Table 6.** Physicochemical Data for **4b–d**



compd	Ar	yield (%)	mp (°C) <sup>a</sup>	formula <sup>b</sup>
<b>4b</b>	3-ClC <sub>6</sub> H <sub>4</sub>	47	146–147	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> Cl <sub>2</sub> OS
<b>4c</b>	4-ClC <sub>6</sub> H <sub>4</sub>	45	151–152	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> Cl <sub>2</sub> OS
<b>4d</b>	3,4-(MeO) <sub>2</sub> C <sub>6</sub> H <sub>3</sub>	35	150–151	C <sub>22</sub> H <sub>27</sub> N <sub>3</sub> O <sub>5</sub> S

<sup>a</sup> All compounds were recrystallized from ethyl acetate. <sup>b</sup> C, H, and N analyses are within ±0.4% of theoretical values.

(0.81 g, 5 mmol) in xylene (50 mL) was refluxed for 5 h and then concentrated *in vacuo*. The residue was chromatographed on silica gel using a CH<sub>2</sub>Cl<sub>2</sub>–AcOEt mixture as an eluent to give **2b** as colorless crystals (0.90 g, 48%). Recrystallization from AcOEt afforded colorless needles: mp 151–152 °C; <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 3.35 (4H, s), 4.58 (4H, s), 7.00 (1H, br s), 7.10–7.45 (8H, m); IR (KBr) 3216 (NH), 1688 cm<sup>-1</sup> (C=O); MS *m/z* 377 (M<sup>+</sup>). Anal. (C<sub>18</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub>Cl<sub>2</sub>) C, H, N.

Other 1,3,5-triazacycloheptane-2,4-diones (**2c,d**) and new 1,3,5-triazacyclooctane-2,4-diones (**3a–d**) were prepared in a manner similar to that described for **2b** from the corresponding ethylenediamine **8** or trimethylenediamine **9**. Physicochemical data are summarized in Table 5.

**1,5-Dibenzyl-4-thioxo-1,3,5-triazacycloheptan-2-one (4a) and 1,5-Dibenzyl-1,3,5-triazacyclooctane-2,4-dithione (5a).** Lawesson's reagent (6.1 g, 15 mmol) was added to a stirred suspension of **2a** (3.1 g, 10 mmol) in toluene (200 mL) at room temperature under N<sub>2</sub> atmosphere. The reaction mixture was heated at reflux for 3 h and evaporated *in vacuo*. The residue was chromatographed on alumina using a PhH–AcOEt mixture as an eluent to give **4a**<sup>20</sup> and **5a**.

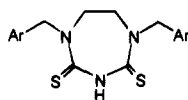
**4a:** 38% yield; mp 139–140 °C; colorless needles (from AcOEt); <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 3.10–3.30 (2H, m), 3.50–3.70 (2H, m), 4.50 (2H, s), 5.15 (2H, s), 7.20–7.40 (10H, br s), 8.20 (1H, br s); IR (KBr) 3232 (NH), 1652 cm<sup>-1</sup> (C=O); MS *m/z* 325 (M<sup>+</sup>). Anal. (C<sub>18</sub>H<sub>19</sub>N<sub>3</sub>OS) C, H, N.

**5a:** 55% yield; mp 165–166 °C; colorless needles (from PhH); <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 3.50 (4H, s), 5.17 (4H, s), 7.33 (10H, m), 9.53 (1H, br s); IR (KBr) 3400 cm<sup>-1</sup> (NH); MS *m/z* 341 (M<sup>+</sup>). Anal. (C<sub>18</sub>H<sub>19</sub>N<sub>3</sub>S<sub>2</sub>) C, H, N.

Other 2-oxo-4-thioxo (**4b–d**) and 2,4-dithione (**5b–d**) compounds were prepared in a manner similar to that described for **4a** and **5a** from the corresponding **2b–d**. Physicochemical data are summarized in Tables 6 and 7.

**1,5-Dibenzyl-4-(methylthio)-1,5,6,7-tetrahydro-2H-1,3,5-triazepin-2-one (6a).** A mixture of **4a** (0.65 g, 2.0 mmol) in DMF (10 mL) and 60% NaH (0.06 g, 2.5 mmol) was stirred for 30 min at room temperature; then methyl iodide (0.28 g, 2.0 mmol) was added to the mixture. The reaction mixture was heated for 2 h at 50 °C and diluted with ice-cold water (15 mL), and the aqueous mixture was extracted with Et<sub>2</sub>O. The extracts were washed with water, dried, and evaporated *in vacuo*. The residue was chromatographed on alumina using a PhH–CH<sub>2</sub>Cl<sub>2</sub> mixture as an eluent to give **6a**: 82% yield; mp 152–153 °C; colorless needles (from isopropyl ether); <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 2.50 (3H, s), 3.15–3.50 (4H, m), 4.58 (2H, s), 4.60 (2H, s), 7.10–7.40 (10H, m); IR (KBr) 1630 cm<sup>-1</sup> (C=O). Anal. (C<sub>18</sub>H<sub>19</sub>N<sub>3</sub>OS) C, H, N.

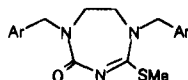
Table 7. Physicochemical Data for 5b-d



compd	Ar	yield (%)	mp (°C) <sup>a</sup>	formula <sup>b</sup>
5b	3-ClC <sub>6</sub> H <sub>4</sub>	45	146–147	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> Cl <sub>2</sub> S <sub>2</sub>
5c	4-ClC <sub>6</sub> H <sub>4</sub>	35	148–149	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> Cl <sub>2</sub> S <sub>2</sub>
5d	3,4-(MeO) <sub>2</sub> C <sub>6</sub> H <sub>3</sub>	60	165–166	C <sub>22</sub> H <sub>27</sub> N <sub>3</sub> O <sub>4</sub> S <sub>2</sub>

<sup>a</sup> All compounds were crystallized from benzene. <sup>b</sup> C, H, and N analyses are within ±0.4% of theoretical values.

Table 8. Physicochemical Data for 6b-d



compd	Ar	yield (%)	mp (°C) <sup>a</sup>	HRMS (m/z) <sup>b</sup>
6b	3-ClC <sub>6</sub> H <sub>4</sub>	82	oil	407.0626 (407.0612)
6c	4-ClC <sub>6</sub> H <sub>4</sub>	95	oil	407.0626 (407.0652)
6d	3,4-(MeO) <sub>2</sub> C <sub>6</sub> H <sub>3</sub>	88	oil	459.1828 (459.1762)

<sup>a</sup> Compounds were purified by column chromatography on alumina. <sup>b</sup> Calcd (found).

Other 4-(methylthio)-2-ones (6b-d) were prepared in a manner similar to that described for 6a from the corresponding 4b-d. Physicochemical data are summarized in Table 8.

**1,5-Dibenzyl-4-(methylthio)-1,5,6,7-tetrahydro-2H-1,3,5-triazepine-2-thione (7a).** A mixture of 5a (1.2 g, 3.5 mmol) in DMF (8 mL) and 60% NaH (0.2 g, 5.0 mmol) was stirred for 20 min at 50 °C; then methyl iodide (0.75 g, 5.3 mmol) was added to the mixture. The reaction mixture was stirred for 3 h at room temperature and diluted with ice-cold water, and the aqueous mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The extracts were washed with water, dried, and evaporated *in vacuo*. The residue was chromatographed on silica gel using an AcOEt-*n*-hexane mixture as an eluent to give 7a: 70% yield; mp 121–122 °C; yellow needles (from diethyl ether); <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ 2.58 (3H, s), 3.28 (2H, m), 3.61 (2H, m), 4.54 (2H, s), 5.26 (2H, s), 7.10–7.60 (10H, m); IR (KBr) 1566 cm<sup>-1</sup> (C=N). Anal. (C<sub>19</sub>H<sub>21</sub>N<sub>3</sub>S<sub>2</sub>) C, H, N.

**Evaluation of Cytotoxicity.** Parent mouse leukemia P388 cells (P388/S) and adriamycin-resistant P388 cells (P388/ADR) were generously provided by the Cancer Chemotherapy Center, Japanese Foundation for Cancer Research, Tokyo, Japan. Cells were passaged weekly in the abdominal cavities of female BALB/c × DAB/2 (CDF<sub>1</sub>) mice (Nippon SLC, Hamamatsu, Japan). To assess the effects on P388/S and P388/ADR cells *in vitro*, cells were suspended at a density of 10<sup>5</sup> cells/mL in RPMI-1640 medium with 10% fetal calf serum, 20 μM β-mercaptoethanol, and 100 μg/mL kanamycin (G-medium). Two hundred microliters of the cell suspension was seeded in each well of 96-well plastic dishes. A test compound was dissolved in dimethyl sulfoxide (DMSO), and the final concentration of DMSO was 0.25% (v/v). The effects of drugs on cell growth were evaluated after consecutive culture for 48 h by MTT assay.<sup>21</sup>

**Evaluation of VBL Accumulation.** Cells (10<sup>6</sup>) were suspended in 1 mL of 20 mM Hepes-buffered G-medium (pH 7.4) and incubated with 4.6 kBq of [<sup>3</sup>H]VBL in the presence or absence of a test compound at 37 °C for 30 min. The final concentration of DMSO was 1%. After the incubation, the cells were chilled on ice and collected by centrifugation (2000 rpm, 5 min) at 2 °C. The cells were washed twice with chilled phosphate-buffered saline (PBS; pH 7.4), and their VBL content was measured by the radioactivity after solubilization with NaOH and neutralization with acetic acid.

**In Vivo Combined Chemotherapy.** Female CD2F<sub>1</sub> mice (5/group) were inoculated intraperitoneally with 1.0 × 10<sup>6</sup> P388/ADR cells/head on day 0 and treated with drugs on days 1–4 and 5–8. Treated drugs were 6c and VBL: 6c was given orally, and VBL was given intraperitoneally 2 h after 6c.

**Materials for Pharmacological Evaluations.** [<sup>3</sup>H]VBL (333 GBq/mmol) was purchased from Moravak Biochemicals. The antitumor drugs used were VBL (Shionogi, Osaka), adriamycin, mitomycin C (Kyowa Hakko Kogyo, Tokyo), and cisplatin (Nippon Kayaku, Tokyo).

**Acknowledgment.** The authors thank Miss. H. Yamada for her skillful technical assistance. This study was supported by a grant from the Ministry of Education, Science, and Culture, Japan, and the Hokkoku Foundation for Cancer Research, Japan.

## References

- Bradely, G.; Juranka, P. F.; Ling, V. Mechanism of Multidrug Resistance. *Biochim. Biophys. Acta* **1988**, *94*, 87–128.
- van der Bliet, A. M.; Borst, P. Multidrug Resistance. *Adv. Cancer Res.* **1989**, *52*, 165–203.
- Tsuruo, T.; Iida, H.; Tsukagoshi, S.; Sakurai, Y. Overcoming of Vincristine Resistance in P388 Leukemia *in Vivo* and *in Vitro* through Enhanced Cytotoxicity of Vincristine and Vinblastine by Verapamil. *Cancer Res.* **1981**, *41*, 1967–1972.
- Tsuruo, T.; Iida, H.; Tsukagoshi, S.; Sakurai, Y. Potentiation of Vincristine and Adriamycin Effects in Human Hemopoietic Tumor Cell Lines by Calcium Antagonists and Calmodulin Inhibitors. *Cancer Res.* **1983**, *43*, 2267–2272.
- Nogae, I.; Kohno, K.; Kikuchi, K.; Kuwano, M.; Akiyama, S.; Kiue, A.; Suzuki, K.; Yoshida, Y.; Cornwell, M. M.; Pastan, I.; Gottesman, M. M. Analysis of Structural Features of Dihydropyridine Analogues Needed to Reverse Multidrug Resistance and to Inhibit Photoaffinity Labeling of P-Glycoprotein. *Biochem. Pharmacol.* **1989**, *38*, 519–527.
- Inaba, M.; Fujikura, R.; Tsukagoshi, S.; Sakurai, Y. Restored *In Vitro* Sensitivity of Adriamycin- and Vincristine-Resistant P388 Leukemia with Reserpine. *Biochem. Pharmacol.* **1981**, *30*, 2191–2194.
- Miyamoto, K.; Wakusawa, S.; Yanaoka, T.; Koshiura, R. Studies on Enhancement of Sensitivity to Vinblastine by Rauwolfia Alkaloids. *Yakugaku Zasshi* **1984**, *104*, 1295–1300.
- Akiyama, S.; Cornwell, M. M.; Kuwano, M.; Pastan, I.; Gottesman, M. M. Most Drugs That Reverse Multidrug Resistance also Inhibit Photoaffinity Labeling of P-Glycoprotein by a Vinblastine Analog. *Mol. Pharmacol.* **1988**, *33*, 144–147.
- Slater, L. M.; Sweet, P.; Stupecky, M.; Gupta, S. Cyclosporin A Reverses Vincristine and Daunorubicin Resistance in Acute Lymphatic Leukemia *in Vitro*. *J. Clin. Invest.* **1986**, *77*, 1405–1408.
- Sawanishi, H.; Wakusawa, S.; Murakami, R.; Miyamoto, K.; Tanaka, K.; Yoshifuji, S. Structure-Activity Relationships of Diamines, Dicarboxamides, and Disulfonamides on Vinblastine Accumulation in P388/ADR Cells. *Chem. Pharm. Bull.* **1994**, *42*, 1459–1462.
- Hagiwara, M.; Wakusawa, S.; Miyamoto, K.; Hidaka, H. Obviation of Drug Resistance and Affinity Purification of P-Glycoprotein by Isoquinolinesulfonamides. *Cancer Lett.* **1991**, *60*, 103–107.
- Hagemann, H. 1,3,5-Triazacycloheptane-2,4-diones. Ger-Offen 2,036,172, 1974.
- Wakusawa, S.; Nakamura, S.; Tajima, K.; Miyamoto, K.; Hagiwara, M.; Hidaka, H. Overcoming of Vinblastine Resistance by Isoquinolinesulfonamide Compounds in Adriamycin-Resistant Leukemia Cells. *Mol. Pharmacol.* **1992**, *41*, 1034–1038.
- Otto, E.; Duerckheimer, W.; Muschaweck, R. Thiatriazepinones. Ger-Offen 2,409,355, 1974.
- Suzuki, H.; Tomida, A.; Nishimura, T. Cytocidal Activity of a Synthetic Isoprenoid, *N*-Solanesyl-*N,N'*-bis(3,4-dimethoxybenzyl)ethylenediamine, and Its Potentiation of Antitumor Drugs against Multidrug-Resistant and Sensitive Cells *in Vitro*. *Jpn. J. Cancer Res.* **1990**, *81*, 298–303.
- Szabo, J. L.; Bruce, W. F. Penicillin Salts of Substituted Alkylene Diamines. U.S. Patent 2,627,491, 1953.
- Binnig, F.; Raschack, M. Antiarrhythmic Alkylene Diamine Derivatives. Ger-Offen 2,438,288, 1974.
- Billmann, J. H.; Meisenheimer, J. L. Hexahydropyridines. III. A Study of 2-Substituted 1,3-Bis(*p*-methoxybenzyl)hexahydropyrimidines and 2-Substituted 1,3-Bis(*p*-chlorobenzyl)hexahydropyrimidines as Transporter Molecules for Tumor Inhibition. *J. Med. Chem.* **1963**, *6*, 682–683.
- Meisenheimer, J. L.; Raya, D. G.; Stapleton, M. C. Synthesis of *N,N'*-Diaryllalkyl 2H-1,2,6-Tetrahydrothiadiazines. *Synth. Commun.* **1989**, *19*, 2229–2236.
- 4a has been reported in the following papers: Dabler, M.; Petter, W. 2,5-*N,N'*-Dibenzyl-2,5,7-triazacycloheptane 1-Thione-6-one,

C<sub>18</sub>H<sub>19</sub>N<sub>3</sub>OS. *Cryst. Struct. Commun.* **1978**, *7*, 321–326. Shanzer, A. Synthesis with metalloid derivatives: Preparation of Heterocycles. *Angew. Chem., Int. Ed. Engl.* **1980**, *19*, 327–328. However, the physicochemical data for **4a** have not been described.

(21) Mosmann, T. Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays. *J. Immunol. Methods* **1983**, *65*, 55–63.

JM950322R